



Synthesis of a new chelating agent derived from phenylenediamine for application in radioimmunotherapy

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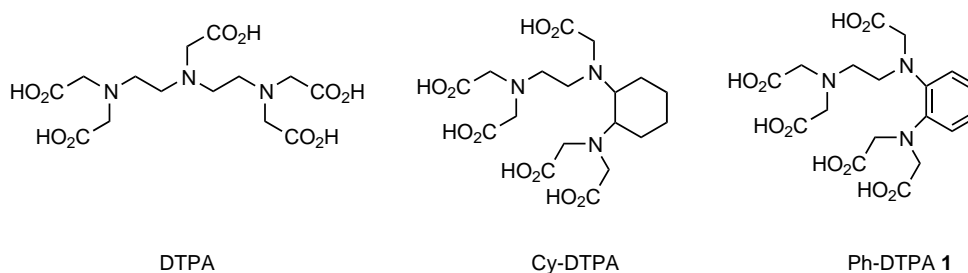
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Abstract—We report the synthesis of a new diethylenetriaminepentaacetic acid (DTPA) analogue, Ph-DTPA, possessing an aromatic rigid skeleton. This precursor of a bifunctional chelating agent was synthesized by elaboration of the intermediate *N*-(*o*-aminophenyl)ethylenediamine and obtained in five steps with an overall yield of 42%. Preliminary complexation studies between Ph-DTPA and ¹⁵³Sm has been carried out. © 2002 Elsevier Science Ltd. All rights reserved.

For many years, complexes combining metals with DTPA and its derivatives have been widely used in magnetic resonance imaging (MRI),^{1–5} radiotherapy^{6–8} and radiodiagnosis.⁹ In particular, the gadolinium–(DTPA) complex is currently used in clinical applications as a contrast agent in MRI.^{10,11} Moreover, many complexes combining lanthanides with DTPA analogues have proved stable enough for use in a physiological medium as radiopharmaceuticals.^{12,13} Since the introduction of this family of chelating agents, developments have been directed essentially at increasing the stability of the resulting complexes.^{14,15} The most promising results have been based on studies to improve the rigidity of the chelating structure. The introduction of a semi-rigid preformed skeleton, which

minimizes the freedom of donor atoms, has had a significant effect on the stability of the metal complexes formed.¹⁶ Notably, the teams of R. D. Hancock and then M. W. Brechbiel have shown that DTPA analogues with a semi-rigid structure of cyclohexyl (Cy-DTPA),^{17,18} piperidinyl (PIP-DTPA) or azaparyl (AZEP-DTPA)¹⁹ type prove considerably more stable in vivo. On this basis, our recent research has been directed at developing stereoisomer chelating agents based on ethylenediaminetetraacetic acid analogues with a cyclopentanic skeleton.²⁰ The synthesis of the DTPA analogue phenyleneethylenetriamine pentaacetic acid (Ph-DTPA) reported here provides a chelating agent even more rigid than Cy-DTPA and with an aromatic skeleton (Scheme 1).



Scheme 1.

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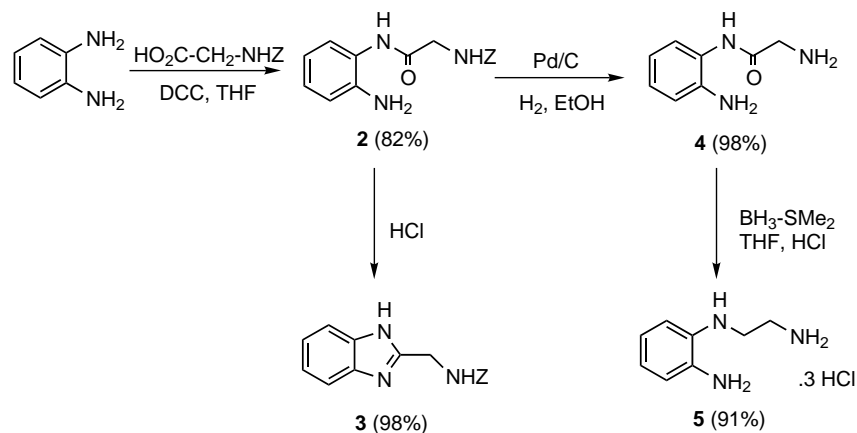
A bifunctional chelating agent is required for the elaboration of bioconjugates suitable for cancer radioimmunotherapy. The functional group commonly used is the *p*-isothiocyanatobenzyl chain, which is generally derived from nitrobenzene that must be introduced during the synthesis of the complexing molecule.^{21,22} The octadentate chelating agent Ph-DTPA has a rigid structure, but is also easy to functionalize directly on the aromatic cycle by means of an isothiocyanate group.

The synthesis of Ph-DTPA **1** was performed in five steps from the starting product, 1,2-phenylenediamine (Scheme 2). The first step, described previously, which consists in peptidic coupling of carbobenzyloxyglycine and the aromatic amine in the presence of dicyclohexylcarbodiimide, yields compound **2**.²³ The synthesis of compound **4** implies the elimination of the Z group. The usual deprotection method for benzyloxycarbonyl (HBr in acetic acid)²⁴ could not be used. In fact, substituted aniline **2** proved unstable in acid medium, yielding quantitatively benzimidazole **3**, a product belonging to a pesticide class.²⁵ The most efficient method for achieving **4**, which provided the best yields (98%) without purification, involved the use of palladium on charcoal as a catalyst under a hydrogen atmosphere. This method proved more suitable than that described by M. R. Bermejo, which uses cyclohexene as hydrogen source, providing yields of no more than 80%.^{26,27}

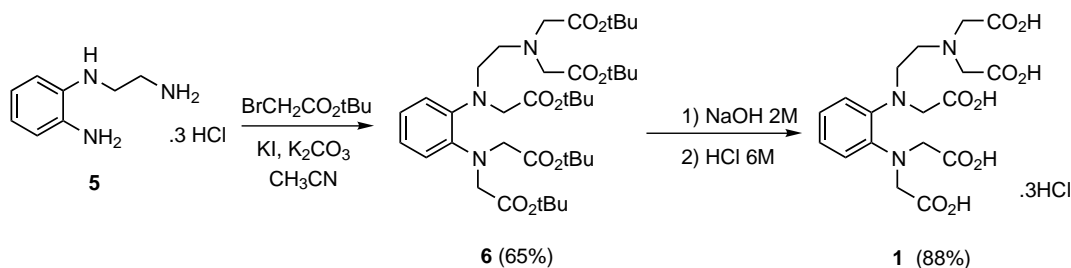
The key intermediate, *N*-(*o*-aminophenyl)ethylenediamine **5**, was obtained with an excellent yield by using the BH₃–Me₂S complex as a reducing agent, followed by treatment with HCl gas.^{28,29} The preparation of triamine **5** was previously reported according to a totally different synthesis strategy, which provided an overall yield of no more than 29%.³⁰ According to the classical alkylation method, penta-*tert*-butyl ester **6** was obtained through the action of excess *tert*-butylbromoacetate in the presence of KI and K₂CO₃ in acetonitrile.³¹ Finally, hydrolysis of the ester functions was performed in basic medium, providing Ph-DTPA **1** with a yield of 88% (Scheme 3).

Preliminary complexation studies between Ph-DTPA and ¹⁵³Sm has been carried out. It shows that the kinetic of complexation of ¹⁵³Sm to **1** is adapted to radioimmunotherapy.³² This was evident from the fact that, on incubating for 45 min at 37°C a radioactive solution containing a 1/1 molar ratio of ¹⁵³Sm/Ph-DTPA, 56% of ¹⁵³Sm chelating agent complex is formed.³³

In conclusion, a rigid DTPA analogue was synthesized with a overall yield of 42% after five steps. Studies of the complexing efficiency of this new chelating agent with radioactive metals suitable for radioimmunotherapy (⁹⁰Y and ¹⁵³Sm) are in progress.



Scheme 2.



Scheme 3.

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References

- Lakshmi, B.; Prabhavathi Devi, A.; Nagarajan, M. *J. Chem. Soc., Perkin Trans. 1* **1997**, 1495–1500.
- Baker, W. C.; Choi, M. J.; Hill, D. C.; Thompson, J. L.; Petillo, P. A. *J. Org. Chem.* **1999**, *64*, 2683–2689.
- Anelli, P. L.; Bertini, I.; Fragai, M.; Lattuada, L.; Luchinat, C.; Parigi, G. *Eur. J. Inorg. Chem.* **2000**, 625–630.
- Zhang, S.; Wu, K.; Sherry, A. D. *Angew. Chem., Int. Ed.* **1999**, *38*, 3192–3194.
- Caravan, P.; Ellison, J. J.; McMurry, T. J.; Lauffer, R. B. *Chem. Rev.* **1999**, *99*, 2293–2352.
- Deal, K. A.; Davis, I. A.; Mirzadeh, S.; Kennel, S. J.; Brechbiel, M. W. *J. Med. Chem.* **1999**, *42*, 2988–2992.
- Volkert, W. A.; Hoffman, T. J. *Chem. Rev.* **1999**, 2269–2292.
- Jang, Y. H.; Blanco, M.; Dasgupta, S.; Keire, D. A.; Shively, J. E.; Goddard, W. A. *J. Am. Chem. Soc.* **1999**, *121*, 6142–6151.
- Anderson, C. J.; Welch, M. J. *Chem. Rev.* **1999**, *99*, 2219–2234.
- Carr, D. W.; Brown, J.; Bydder, G. M. *Lancet* **1984**, *1*, 484–489.
- Lauffer, R. B. *Chem. Rev.* **1987**, *87*, 901–927.
- Liu, S.; Edwards, S. *Bioconjugate Chem.* **2001**, *12*, 7–34.
- Wu, C.; Kobayashi, H.; Sun, B.; Yoo, T. M.; Paik, C. H.; Gansow, O. A.; Carrasquillo, J. A.; Pastan, I.; Brechbiel, M. W. *Bioorg. Med. Chem.* **1997**, *5*, 1925–1934.
- Grote, C. W.; Kim, D. J.; Rapoport, H. *J. Org. Chem.* **1995**, *60*, 6987–6997.
- Williams, M. A.; Rapoport, H. *J. Org. Chem.* **1993**, *58*, 1151–1158.
- Fossheim, R.; Dugstad, H.; Dahl, S. G. *J. Med. Chem.* **1991**, *34*, 819–826.
- De Sousa, A. S.; Croft, G. J. B.; Wagner, C. A.; Michael, J. P.; Hancock, R. D. *Inorg. Chem.* **1991**, *30*, 3525–3529.
- McMurry, T. J.; Pippin, C. G.; Wu, C.; Deal, K. A.; Brechbiel, M. W.; Mirzadeh, S.; Gansow, O. A. *J. Med. Chem.* **1998**, *41*, 3546–3549.
- Chong, H.-S.; Garmestani, K.; Bryant, H.; Brechbiel, M. W. *J. Org. Chem.* **2001**, *66*, 7745–7750.
- Gouin, S. G.; Gestin, J.-F.; Joly, K.; Loussouarn, A.; Reliquet, A.; Meslin, J.-C.; Deniaud, D. *Tetrahedron* **2002**, *58*, 1131–1136.
- Gestin, J. F.; Loussouarn, A.; Bardies, M.; Gautherot, E.; Gruaz-Guyon, A.; Saï Maurel, C.; Bardet, J.; Curtet, C.; Chatal, J. F.; Faivre Chauvet, A. *J. Nucl. Med.* **2001**, *42*, 146–153.
- Brechbiel, M. W.; Gansow, O. A. *J. Chem. Soc., Perkin Trans. 1* **1992**, 1173–1178.
- Hoyos, O. L.; Bermejo, M. R.; Fondo, M.; Garcia-Deibe, A.; Gonzalez, A. M.; Maneiro, M.; Pedrido, R. *J. Chem. Soc., Dalton Trans. 1* **2000**, 3122–3127.
- Ben-Ishai, D.; Berger, A. *J. Org. Chem.* **1952**, *17*, 1564–1566.
- Maekawa, K.; Taniguchi, E.; Kuwano, E.; Shuto, Y. *Environ. Qual. Saf. Suppl.* **1975**, *3*, 748–753.
- Bermejo, M. R.; Gonzalez, A. M.; Fondo, M.; Garcia-Deibe, A.; Maneiro, M.; Sanmartin, J.; Hoyos, O. L.; Watkinson, M. *New J. Chem.* **2000**, *24*, 235–241.
- Jackson, A. E.; Johnstone, R. A. W. *Synthesis* **1976**, 685–687.
- Moreau, P.; Tinkl, M.; Tsukazaki, M.; Bury, P. S.; Griffen, E. J.; Snieckus, V.; Maharajh, R. B.; Kwok, C. S.; Somayaji, V. V.; Peng, Z.; Sykes, T. R.; Noujaim, A. A. *Synthesis* **1997**, 1010–1012.
- Achmatowicz, M.; Jurczak, J. *Tetrahedron: Asymmetry* **2001**, *12*, 487–495.
- Linsker, F.; Evans, R. L. *J. Org. Chem.* **1945**, *10*, 283–285.
- Galaup, C.; Couchet, J.-M.; Picard, C.; Tisnès, P. *Tetrahedron Lett.* **2001**, *42*, 6275–6278.
- Li, W. P.; Ma, D. S.; Higginbotham, C.; Hoffman, T.; Ketring, A. R.; Cutler, C. S.; Jurisson, S. S. *Nucl. Med. Biol.* **2001**, *28*, 145–154.
- Complexation studies with ^{153}Sm .** Stock solution of Ph-DTPA ($0.11 \text{ mg}\cdot\text{mL}^{-1}$) was prepared in 0.1 M sodium acetate buffer (pH 5.6). To form ^{153}Sm chelating agent complex, $5 \mu\text{L}$ (3.70 nmol) of ^{153}Sm stock solution ($140.5 \text{ mCi}\cdot\text{mL}^{-1}$) was added to 1 equiv. of Ph-DTPA. The solution was made up to $500 \mu\text{L}$ (final pH of the solution was 5.6, with a final ^{153}Sm concentration of $7.5 \text{ nmol}\cdot\text{mL}^{-1}$) and incubated at 37°C for 45 min. The complexation was measured on a phosphoimager 445SI after thin-layer chromatography on cellulose plates (Merck 5552/0025) by elution with 0.1 M sodium acetate (pH 5.6)/methanol (2/1).